INVITRO ANTI-INFLAMMATORY ACTIVITY OF PARKIA BIGLOBOSA FRUIT BARK EXTRACT

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ABSTRACT- The invitro anti-inflammatory activity of various solvent fractions of Parkia biglobosa fruit bark was investigated using human red blood cell membrane stabilization, heat-induced hemolysis and protein denaturation methods. All the extracts of P. biglobosa fruit bark showed a concentration dependent increase in anti-inflammatory activity. The anti-inflammatory activity of the crude extract (60.8%, 58.3%, 78.2%) and last remaining aqueous extract (61.1%, 54.1%, 77.2%) have the maximum membrane stabilization, protection against hemolysis and albumin denaturation respectively which was comparable to Diclofenac sodium (61.4%, 60.6%, 100%) at 400µg/ml concentration. This study suggests that P. biglobosa fruit bark posses enough potential to reduce inflammation, hence directs the importance of further research and development of novel anti-inflammatory agents.

Key words- Invitro anti-inflammatory, fruit bark, HRBC membrane stabilization, hemolysis, protection

1.0 INTRODUCTION

Inflammation is the reaction of living tissue to injury, infection or irritation. It involves complex array of enzyme activation, mediator release, fluid extravasations, cell migration, tissue breakdown and repair [1] which is aimed at host defence and usually activated in most disease condition. Drugs which are in use presently for the management of pain and inflammatory conditions are either narcotics e.g. opioids or non-narcotics e.g. salicylates and corticosteroids e.g. hydrocortisone. Due to risk of adverse effects encountered with the use of synthetic anti-inflammatory drugs, natural products derived from plants may offer an alternative source of anti-inflammatory agent. The use of traditional medicine and medicinal plants in most developing countries, as a normative basis for the maintenance of good health, has been widely observed [2].

Parkia biglobosa is widely acclaimed by the Hausa communities of northern Nigeria for the treatment of diseases such as malaria, diabetes mellitus and pains. The stem barks is boiled in water and taken as a decoction for the treatment of malaria, inflammatory diseases and infections to diarrhea [3]. The bark soaked in ethanol is also used in some communities for anti-diarroheal properties and as effective anti-snake venoms that protects against neurotoxic, haemotoxic and cytotoxic effects of poisonous snakes [4]. Also, the leaves, fruits and seeds of P. biglobosa have also been used to manage various diseases [5].

2.0 MATERIALS AND METHODS

2.1 Plant material collection and preparation

The fruit back of P. biglobosa were collected in April, 2014 from Bimin Yauri Town, Ngaski Local Government Area, Kebbi State, Nigeria. The plant was identified by their local names and later authenticated by the herbarium at Department of Biological Sciences, Kebbi State University of Science and Technology, Aliero with a voucher specimen (No 281). Fresh fruit back were air-dried and grinded into coarse powder using mortar and pestle.

2.2 Extraction of the plant material

Crude extraction was carried out according to the method of Edeoga et al [6] with slight modifications. One kilogram (1.0 kg) of the fruit bark powder was extracted with hydromethanolic solvent (methanol (70): water (30)) for 72 hours. This was then filtered and evaporated to dryness in an oven at 45°C to give hydromethanolic crude extract (346g).

2.3 Phytochemical analysis

Preliminary phytochemical studies were conducted as per standard procedures described by Trease and Evan [7] and Harbone [8]

2.4 Solvent Partitioning of the Methanolic Extract

Hydromethanolic extract (100g) was redissolved in distilled water (200 mL) in a separating funnel and partitioned with n-hexane (4 x 200 mL). The resulting n-hexane phase was concentrated to dryness and known as hexane fraction which was stored in a freezer in an air-tight container. The resulting aqueous phase was further partitioned with chloroform,
ethyl acetate and butanol respectively. The last remaining aqueous fraction was then evaporated to dryness and kept for further use in the freezer in an air-tight container [6].

2.5 Invitro anti inflammatory studies

2.5.1 Preparation of Red Blood cells (RBCs) suspension

Fresh whole human blood (5mL) was collected and transferred to the centrifuged tubes containing Heparin or EDTA or Sodium citrate to prevent clotting. The tubes were centrifuged at 3000 rpm for 10 min and were washed three times with equal volume of normal saline. The volume of the blood was measured and reconstituted as 10% v/v suspension with normal saline [9].

2.5.2 The human red blood cell (HRBC) membrane stabilization method.

The reaction mixture consist of 1.0ml of test sample of different concentration (50-400µg/ml) in normal saline and 0.5 ml of 10% HRBC suspension, 1 ml of 0.2 M phosphate buffer, 1 ml hypo saline were incubated at 37°C for 30 min and centrifuged at 3000 rpm for 20 min and the hemoglobin content of the supernatant solution was estimated spectrophotometrically at 560 nm. Diclofenac sodium was used as standard and a control was prepared without extracts [10]. The percentage of hemolysis and membrane stabilization or protection was calculated by using the following formula;

\[
\% \text{ Hemolysis} = \left( \frac{\text{Optical density of test sample}}{\text{Optical density of control}} \right) \times 100
\]

\[
\% \text{ Protection} = 100 - \left( \frac{\text{Optical density of test sample}}{\text{Optical density of control}} \right) \times 100
\]

2.5.3 Heat induced human red blood cell (HRBC) membrane stabilization method

The reaction mixture in heat induced hemolysis consists of 1.0 ml of test sample of different concentrations (50μg – 400 µg), 0.2 ml of 0.05 % BSA and 1.8 ml of 0.2 M phosphate buffered saline (pH 6.4). The mixtures were incubated at 37°C for 15 minutes and then heated at 70°C for 5 minutes. After cooling, the absorbance was measured spectrophotometrically at 660nm against a blank. Diclofenac sodium was used as standard drug and the percentage inhibition of protein denaturation was calculated by using the following formula;

\[
\% \text{ Inhibition} = 100 \times \left( 1 - \frac{V_t}{V_c} \right)
\]

Where, Vt = absorbance of test sample; Vc = absorbance of control.

3.0 RESULTS AND DISCUSSION

3.1 Percentage Yield and Phytochemistry

The percentage yield of 1000g of P. biglobosa fruit bark using hydroethanolic extraction was 34.6% while fractionation of 100g of hydroethanolic extract of P. biglobosa fruit bark obtained percentage yield as presented in Table 1. Phytochemical screening which revealed the present of secondary metabolites as shown below in Table 2.

Table 1: Percentage yield of P. biglobosa fruit bark (100g)

<table>
<thead>
<tr>
<th>Fraction</th>
<th>Percentage yield (%)</th>
</tr>
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<tbody>
<tr>
<td>Hexane</td>
<td>28.6</td>
</tr>
<tr>
<td>Chloroform</td>
<td>23.5</td>
</tr>
<tr>
<td>Ethyl acetate</td>
<td>21.2</td>
</tr>
<tr>
<td>Butanol</td>
<td>4.4</td>
</tr>
<tr>
<td>Last remaining aqueous fraction</td>
<td>14.8</td>
</tr>
</tbody>
</table>

Table 2 : Preliminary Phytochemical screening of P. biglobosa fruit bark

<table>
<thead>
<tr>
<th>Phytochemicals</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkaloids</td>
<td>+</td>
</tr>
<tr>
<td>Tannins</td>
<td>++</td>
</tr>
<tr>
<td>Saponins</td>
<td>ND</td>
</tr>
<tr>
<td>Flavonoids</td>
<td>+++</td>
</tr>
<tr>
<td>Steroids</td>
<td>++</td>
</tr>
</tbody>
</table>
Cardiac glycosides | +
---|---
Reducing sugar | ++

(+): trace amount, (++) moderately present, (+++) highly present, (ND): Not Detected

3.2 Membrane stabilization activity of P. biglobosa fruit bark

Lysosomal enzymes released during inflammation produce a variety of disorders. The extra cellular activity of these enzymes is said to be related to acute or chronic inflammation. The non steroidal drugs act either by inhibiting these lysosomal enzymes or by stabilizing the lysosomal membrane [13]. The invitro anti-inflammatory activity of P. biglobosa fruit bark fractions were assessed by HRBC membrane stabilization method (Figure 1). Crude extract and fractions of P. biglobosa fruit bark exhibited varying degree of anti-inflammatory activity which was comparable to the standard drug Diclofenac sodium. The result showed a concentration dependent increase in percentage protection of the membrane in all the tested fractions.

Figure 1: Membrane Stabilization Effect of Parkia biglobosa fruit bark

Since the human red blood cell membrane is analogous to the lysosomal membrane [14], its stabilization indicates the stabilization of lysosomal membranes. Stabilization of the membranes of these cells inhibits lysis and subsequent release of the cytoplasmic contents which in turn limits the tissue damage and exacerbation of the inflammatory response [15]. In this study, P. biglobosa fruit bark was able to confer a significant protection of HRBC membrane when compared to Diclofenac sodium.

3.3 Inhibition of Heat-induced hemolysis

Protein denaturation is the process in which protein lose their secondary and tertiary structure by application of external stress or compounds such as strong acid or base, organic solvent or heat [17]. Denaturation of protein is a well documented cause of inflammation. In the present study, P. biglobosa fruit bark was effective in inhibiting albumin denaturation (Figure 3). Maximum inhibition of 78.2% and 77.2% was observed in hydromethanolic extract and last remain aqueous fraction respectively at a concentration of 400µg/ml. On the other hand, Diclofenac sodium, the standard anti-inflammatory drug showed a maximum inhibition of 100%. The increased percentage of inhibition of albumin denaturation expresses the anti-inflammatory efficacy of P. biglobosa fruit bark.

Figure 3: Effect of Parkia biglobosa Fruit Bark on Albumin Denaturation
4.0 CONCLUSION

The present study indicates that the different solvent extract of *P. biglobosa* fruit bark possess anti-inflammatory property. This activity may be due to presence of phenolic compounds such as flavonoids and tannins. This invitro study is a preliminary evaluation of anti-inflammatory activity of *Parkia biglobosa* fruit bark and validates the folkloric use of this plant in the treatment of inflammatory conditions. Further research work to evaluate in vivo activity on animals, isolation and characterization of its bioactive components is recommended.

REFERENCES


